

A New Bio-Inorganic Nanocomposite Membrane for Glucose-Modulated Release of Insulin

Claudia R. Gordijo, Adam J. Shuhendler and Xiao Yu Wu*

Leslie L. Dan Faculty of Pharmacy, University of Toronto, Toronto, ON, Canada.

xy.wu@utoronto.ca



UNIVERSITY OF TORONTO

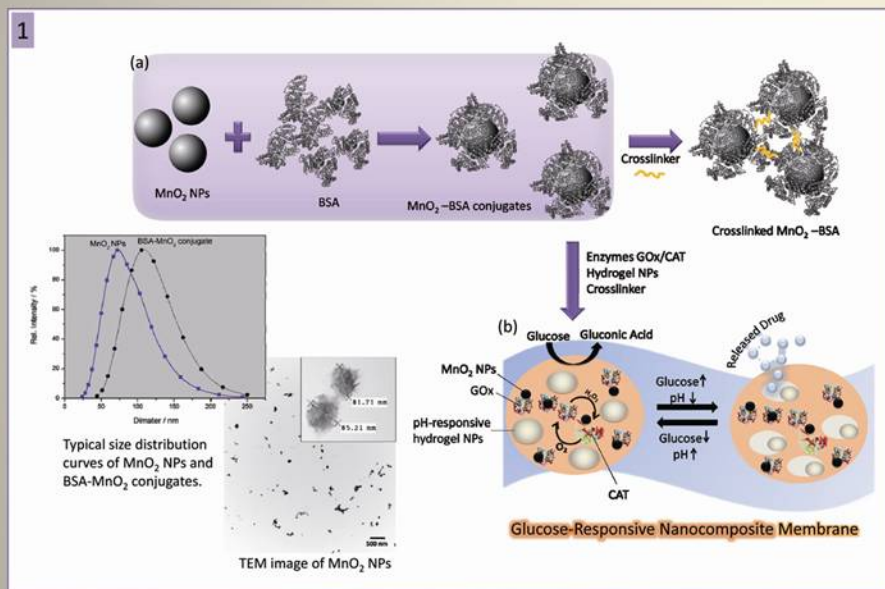
Introduction

Diabetes mellitus is a major public health problem that affects more 246 million people worldwide. It is a disorder in glucose regulation, characterized by the accumulation of glucose in the blood due to the inability of the pancreas to secrete insulin or the body to respond to insulin. The conventional way of controlling glycemia in insulin-dependent diabetic patients is the frequent self-administration of insulin injections, which often results in hypoglycemia along with poor patient compliance. A more effective approach to delivering insulin in direct response to blood glucose levels (mimicking a healthy human pancreas) is thus highly desirable. For this propose, smart nanomaterials have been investigated for glucose-modulated insulin delivery. Our group has been developing glucose-responsive nanocomposite membranes able to control the permeation of insulin in response to glucose concentration. Our approach involves the incorporation of pH-sensitive hydrogel NPs and the enzyme glucose oxidase (GOx) into a polymeric matrix. The membrane acts as a glucose sensor by the action of GOx, which catalyzes the oxidation of glucose to gluconic acid. This reaction creates a low pH microenvironment in the membrane, causing the hydrogel NPs to shrink, leading to the formation of an interconnected porous framework that increases insulin permeation across the membrane (Fig. 1b). In this system, catalase (CAT) is normally co-immobilized with GOx in order to quench the H₂O₂ produced during GOx turn over cycles and replenish ½ of the O₂ consumed by GOx reaction. Still, several factors can lead to GOx inactivation and consequently to the failure of regulated insulin release. Besides the accumulation of hydrogen peroxide which leads to deleterious effects on GOx activity, the hydrophobic nature of the polymeric matrices utilized until now and the harsh conditions applied for the preparation of membranes can also contribute to GOx inactivation.

In order to improve GOx stability and optimize the membrane response to glucose concentration a hydrophilic membrane matrix, an effective H₂O₂ scavenger and mild conditions for membrane preparation are highly desirable. Herein, we propose the application of a new bio-inorganic hybrid material composed of crosslinked bovine serum albumin (BSA) and MnO₂ nanoparticles as a matrix for a glucose-responsive membrane. MnO₂ NPs shows high reactivity towards H₂O₂ while crosslinked BSA presents several advantages over other polymeric matrices previously applied; e.g., high biocompatibility; GOx/CAT immobilization can be processed in an aqueous medium quickly in one-step during membrane preparation. In this way, the combination of crosslinked BSA with MnO₂ NPs could greatly improve GOx stability and consequently lead to a better response to glucose concentration for the controlled release of insulin.

1 – Glucose-Responsive Nacomposite Membrane

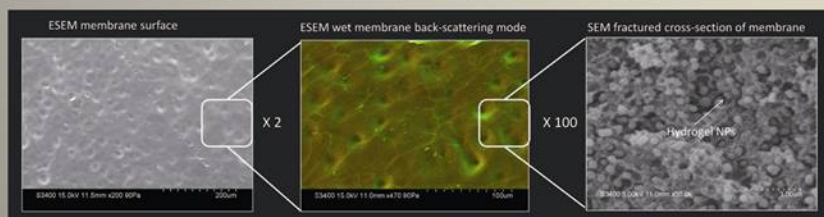
Membrane preparation: multifunctional MnO₂ NPs were conjugated with bovine serum albumin (MnO₂ – BSA) and enzymes GOx and CAT. A membrane was prepared by crosslinking these biomolecule-MnO₂ conjugates with glutaraldehyde in the presence of pH-responsive poly(N-isopropylacrylamide-co-methacrylic acid) (PNIPAM/MAA) hydrogel nanoparticles.



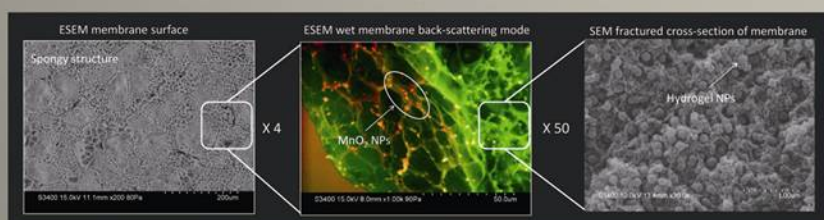
2 – Effect of MnO₂ NPs on Morphology of the Membrane

The incorporation of MnO₂ NPs into the membrane formulation led to different membrane morphology. Unlike the membrane made with crosslinked BSA (Fig. 2a), SEM images of the nanohybrid membranes (made with crosslinked MnO₂ – BSA) revealed a complex sponge-like structure with MnO₂ and hydrogel nanoparticles homogeneously distributed in the thin cavity wall of the base membrane (Fig. 2b).

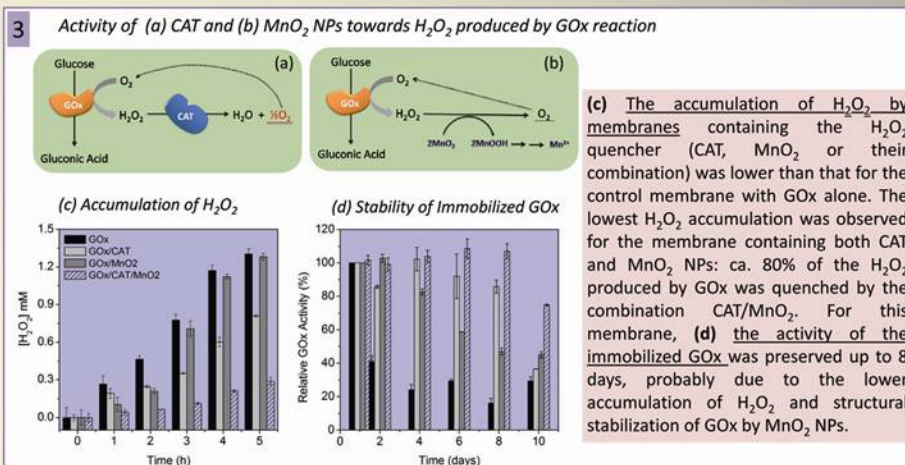
a) Membrane without MnO₂ NPs: crosslinked BSA + enzymes + hydrogel nanoparticles



b) Membrane with MnO₂ NPs: crosslinked BSA-MnO₂ + enzymes + hydrogel nanoparticles



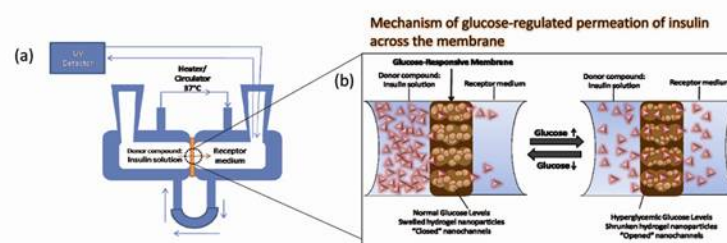
3 – Quenching of H₂O₂ by CAT/MnO₂ NPs and Stability of Immobilized GOx



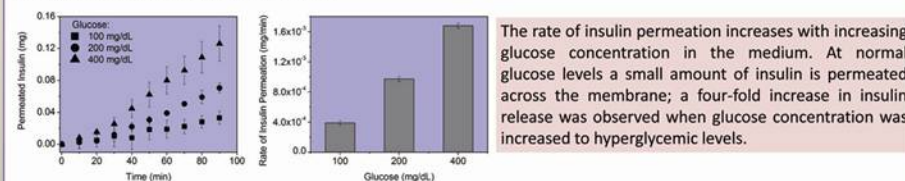
(c) The accumulation of H₂O₂ by membranes containing the H₂O₂ quencher (CAT, MnO₂ or their combination) was lower than that for the control membrane with GOx alone. The lowest H₂O₂ accumulation was observed for the membrane containing both CAT and MnO₂ NPs: ca. 80% of the H₂O₂ produced by GOx was quenched by the combination CAT/MnO₂. For this membrane, (d) the activity of the immobilized GOx was preserved up to 8 days, probably due to the lower accumulation of H₂O₂ and structural stabilization of GOx by MnO₂ NPs.

4 – Insulin Permeation in Response to Glucose Concentration

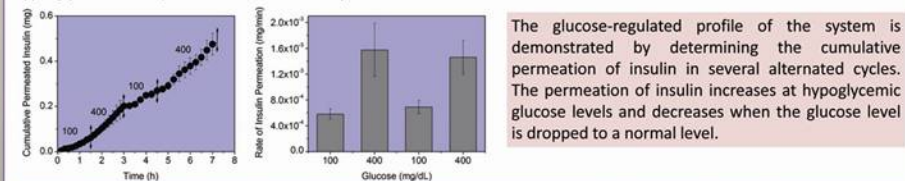
The *in vitro* release of insulin was measured by using a side-by-side diffusion cell system coupled to an UV spectrophotometer (a). The glucose concentration in the release medium was alternated between hypoglycemic and hyperglycemic levels (100 – 400 mg/dL) in order to simulate glucose fluctuations in diabetic patients.



(c) Profile of insulin permeated across the nanocomposite membrane containing GOx and both H₂O₂ quenchers (MnO₂ NPs and CAT) in different glucose concentrations.



(d) Cumulative permeated insulin in response to abrupt changes in glucose concentration (between normal and hyperglycemic levels) in several consecutive cycles.



Conclusions

For the first time a bio-inorganic nanocomposite membrane was prepared for the glucose-modulated release of insulin. The membrane acts as a glucose sensor and insulin release attenuator. The combination of MnO₂ nanoparticles with the enzyme CAT successfully quenched up to 80% of the undesirable hydrogen peroxide produced during GOx cycles, which directly reflected on the improvement of the long term GOx stability in this membrane. The glucose-responsiveness of the system allows for regulated insulin permeation across the membrane in response to normal or hyperglycemic glucose concentrations.

References

K. Zhang, X. Y. Wu, *Journal of Controlled Release*, 2002, 80, 169. / K. Zhang, X. Y. Wu, *Biomaterials*, 2004, 25, 5281.

The authors gratefully acknowledge the Natural Sciences and Engineering Research Council of Canada & BioDiscovery Toronto for the financial support, Dr Hui Yu Huang for the initial studies with crosslinked BSA membranes.